

1 Polymer-coated sample cup for quantitative analysis of semi-volatile
2 phthalates in polymeric materials by thermal desorption-gas
3 chromatography-mass spectrometry

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12

13 Abstract

14 A new “polymer-coated” sample cup useful for the analysis of phthalates
15 in polymeric materials by thermal desorption (TD)-GC/MS using a temperature
16 programmable furnace type pyrolyzer as a TD device was developed to
17 suppress the emission of semi-volatile phthalates such as dimethyl phthalate
18 (DMP) and diethyl phthalate (DEP) during the measurements. The inner
19 surface of a sample cup was coated by polymers which act as a sorbent for the
20 phthalates. Three polymers, polyvinyl chloride, polystyrene and poly (methyl
21 methacrylate), were chosen as the coating polymers. A mixture of ten
22 phthalates including DMP and DEP was used as the test sample to estimate the
23 performance of the sample cups. When a conventional sample cup without
24 any polymer coating was used, 90 and 50% reductions in the peak areas of
25 DMP and DEP were respectively observed at the waiting time of 200 min. On
26 the contrary, no reduction of peak area of DMP and DEP during the same

27 waiting time was observed with any one of the three coating polymers at the
28 proper polymer film thickness. These results suggest that the polymer-coated
29 sample cup suppresses the emission of semi-volatile phthalates and is
30 effective for the analysis of phthalates containing DMP and DEP by TD-GC/MS.

31

32 Keywords

33 dimethyl phthalate, diethyl phthalate, thermal desorption-GC/MS, pyrolyzer,
34 polymer-coated sample cup

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42 1. Introduction

43 Phthalic acid esters, known as phthalates, are used as additives to
44 modify physical properties of polymeric materials. Some phthalates such as
45 dimethyl phthalate (DMP), diethyl phthalate (DEP), dibutyl phthalate (DBP),
46 butyl benzyl phthalate (BBP), di-(2-ethylhexyl) phthalate (DEHP) and di-n-octyl
47 phthalate (DNOP) are widely used as plasticizers to increase flexibility and
48 durability of various polymeric materials such as cellulose esters and polyvinyl
49 chloride (PVC) [1, 2]. Until now, phthalate-containing thermoplastic materials
50 have been widely used in applications ranging from food containers, food
51 wrappers, housewares, toys, electronic devices, and cosmetic products, to
52 medical devices. Because phthalate additives are not chemically bound to the
53 polymer matrix, some of them are released slowly into the external
54 environment from the materials. In addition, phthalates have attracted much
55 public attention because of their carcinogenic and estrogenic properties [3]. As

56 a result, they ubiquitously exist as hazardous chemicals in a variety of
57 substances including aerosols, indoor and outdoor air, dust, food, and even in
58 human urine and blood [4-6]. In this context, some regulations on the use of
59 phthalates have been announced by several government agencies [7-9].

60 Determination of phthalates is mostly carried out by gas chromatography
61 coupled with mass spectrometry (GC/MS). In the case of phthalates in water,
62 air, and particulates, isolation and/or enrichment of the phthalates from a
63 sample material has usually been done by using liquid-liquid extraction [10]
64 and solid-phase microextraction [11, 12], prior to the GC/MS measurements.
65 On the other hand, for the determination of phthalates in solid samples such as
66 polymer materials, sample pretreatment using solvent extraction has been
67 generally carried out prior to the GC/MS measurements [13-15]. However,
68 solvent extraction requires tedious and time-consuming procedures along with
69 use of large amounts of harmful organic solvents. In contrast, phthalates in
70 polymer materials can be directly analyzed in the solid state by thermal
71 desorption (TD)-GC/MS without any pretreatment of samples as described
72 previously [16]. In brief, a several hundred μg of a solid sample is simply
73 placed in a sample cup which falls freely into the pre-heated micro-furnace of
74 the pyrolyzer, and then the sample is heated under the computer-programmed
75 temperature conditions. Since this technique requires minimal sample
76 pre-treatment procedures, Accordingly, TD-GC/MS has been recently
77 adopted as an official method of analysis for phthalates in polymeric materials
78 [17].

79 However, the adopted official method was established targeting only six
80 less-volatile phthalates, DBP, BBP, DEHP, diisodecyl phthalate (DIDP),
81 diisononyl phthalate (DINP) and DNOP, that are restricted by the Consumer
82 Product Safety Improvement Act and other similar legislation [7-9], although
83 other phthalates such as DMP, DEP, di-n-propyl phthalate (DPRP) and
84 diisobutyl phthalate (DIBP) are also environmental contaminants and human

85 health hazards. When applying TD-GC/MS to the analysis of semi-volatile
86 phthalates with a high vapor pressure such as DMP and DEP, accurate
87 determination becomes difficult because of evaporation of such phthalates
88 from the sample cup during the sample preparation procedures. For example,
89 the vapor pressure of DMP at 25°C, 0.26 Pa, is 50 times higher than that of
90 DBP at 25°C, 4.7×10^{-3} Pa [18], and reliable quantitative analysis of such high
91 vapor pressure phthalates is almost impossible.

92 In the present study, the inner surface of a sample cup was coated with
93 polymers which act as a sorbent for phthalates including DMP and DEP to
94 reduce the effect of evaporation of phthalates on their quantitative analysis by
95 TD-GC/MS. PVC, polystyrene (PS) and poly (methyl methacrylate) (PMMA)
96 were examined as the sorbent polymer coating, and attention was paid to their
97 effects on the evaporation reduction for phthalates and on background signals
98 in chromatograms caused by the thermal decomposition of sorbent polymers at
99 elevated temperature.

100

101 2. Experimental

102 2.1 Samples

103 Phthalate compounds used in this work were analytical grade supplied by
104 Tokyo Chemical Industry Co., Ltd. (Tokyo, Japan) and are listed in Table 1
105 together with their abbreviations, boiling point [19], and vapor pressure at 25°C
106 [18]. 200 µg of each phthalate was added into 10 mL dichloromethane (DCM)
107 and thoroughly dissolved. This was used as the stock mixture for the analysis
108 experiment. An aliquot (5 µL) of the dichloromethane solution that contained
109 20 ppm (100 ng) of each phthalate was put in a sample cup using a micro
110 syringe and subjected to the TD-GC/MS analysis.

111 2.2 Polymer-coated sample cup

112 Figure 1 shows a cross-sectional view of a sample cup coated with a
113 sorbent polymer film. The sample cup is made of Pyrex glass (model Eco-cup
114 G, Frontier Laboratories Ltd., Koriyama, Japan) and has the dimensions of:
115 height, 8 mm; outer diameter, 4 mm; inner diameter (i.d.), 3 mm; wall thickness,
116 0.5 mm; and volume, about 50 μL . The sorbent polymers PVC, PS and PMMA
117 were produced industrially. Additives and semi-volatile compounds (SVOCs)
118 in these polymers were removed by Soxhlet extraction for 12 h using hexane as
119 a solvent prior to the formation of the sorbent polymer film inside the sample
120 cup. Tetrahydrofuran (THF) was used for PVC, and DCM was used for PS and
121 PMMA as a solvent to make solutions for the film coating. 20 μL of a polymer
122 solution was put in the sample cup using a micro syringe and a polymer film
123 was formed on the inner surface of the sample cup by evaporating the solvent
124 while heating in an electrical furnace at 50°C for 5 min, and at 200°C for 10 min
125 as a final preparation procedure. The polymer film was not formed uniformly
126 and became thicker near the bottom of the sample cup. The average
127 thickness of the film can be tuned from 0.2 to 4.8 μm by changing the
128 concentration of polymers from 0.5 to 12 $\mu\text{g}/\mu\text{L}$.

129 2.3 TD-GC/MS measurements

130 Figure 2 shows a schematic diagram of the TD-GC/MS system used in
131 this study. A temperature-programmable furnace type pyrolyzer (Multi-Shot
132 Pyrolyzer, EGA/PY-3030D, Frontier Laboratories Ltd.) was directly coupled
133 with a quadrupole GC/MS (Shimadzu QP-2010 Plus or Agilent 5975) equipped
134 with a metal capillary separation column (Ultra ALLOY⁺-5, 30 m \times 0.25 mm i.d.,
135 coated with 5% diphenyl 95% dimethylpolysiloxane in 0.25 μm film thickness,
136 Frontier Laboratories Ltd.). The pyrolyzer was also coupled with an
137 automated sample introduction system (Auto-Shot Sampler, AS-1020E,
138 Frontier Laboratories Ltd.). Sample cups were placed at the waiting position
139 on the top of the Auto-Shot Sampler and were held at room temperature (ca.
140 25°C) before TD-GC/MS measurements. Then, the sample cup was dropped to

141 the heating position at the center of the pyrolyzer furnace which was
142 temperature-controlled. The furnace temperature was increased from 100 to
143 320°C at a rate of 20°C/min, and held at 320°C for 5 min. According to the
144 authors' previous study on evolved gas analysis (EGA) [20] of phthalates in
145 PVC [16], these experimental conditions are sufficient to attain thermal
146 desorption of phthalates from the sample. Gases evolved from the sample
147 were transported to the GC injection port under a 24 mL/min He carrier gas flow
148 that was introduced into the metal capillary separation column with a split ratio
149 of 1:20. The column temperature was programmed from 50°C to 200°C at a
150 rate of 40°C/min and then to 280°C at a rate of 5°C/min. The components
151 separated by the column were finally detected by MS with an electron
152 ionization source and identified by their retention times and mass spectra
153 followed by a mass spectral library search.

154

155 3. Results and discussion

156 3.1 Applicability of a conventional sample cup without polymer coating

157 Figure 3 shows chromatograms of the mixture of phthalates measured by
158 TD-GC/MS using a sample cup without any polymer coating. Figure 3(a) was
159 obtained by the immediate measurement after putting the sample solution into
160 the sample cup, while Fig. 3(b) was obtained after holding the sample cup at
161 the waiting position for 200 min. By comparing both chromatograms in Fig. 3,
162 it can be easily recognized that the intensities of peaks 1 and 2 corresponding
163 to DMP and DEP respectively decrease significantly in Fig. 3(b). On the other
164 hand, peaks 3-10 corresponding to less-volatile phthalates do not show any
165 noticeable decrease in the peak area due to their vapor pressures being
166 sufficiently low at 25°C.

167 Figure 4 shows the dependency of the peak areas of phthalates on the

168 waiting time at 25°C. It is seen that the peak areas of DMP and DEP decrease
169 remarkably as the waiting time increases, and 90 and 50% reductions in the
170 peak areas are respectively seen for DMP and DEP at the waiting time of 200
171 min. These results suggest that DMP and DEP were emitted to the ambient
172 atmosphere during the waiting time; therefore, a reliable calibration curve for
173 the quantitative analysis of semi-volatile phthalates having high vapor
174 pressures cannot be obtained by TD-GC/MS using the conventional sample cup
175 without any polymer coating. Accordingly, it can be said that additional
176 procedures are necessary to determine semi-volatile phthalates by TD-GC/MS.

177

178 3.2 Analysis of semi-volatile phthalates using a polymer-coated sample cup

179 To reduce the emission of semi-volatile phthalates during the waiting
180 period before TD-GC/MS measurements, the effect from coating the inner
181 surface of a sample cup with polymers was examined, with the expectation that
182 the coated polymer functions as a sorbent for semi-volatile phthalates. PVC,
183 PS and PMMA were separately used as the coating polymer in this study for the
184 following reasons. PVC is the most popular polymer containing large amount
185 of phthalates as plasticizers. When heated at high temperatures, PS and
186 PMMA degrade to a few kinds of pyrolyzates which might interfere with the
187 measurements of phthalates, according to their reported pyrograms [21].
188 Furthermore, all these polymers are soluble in a variety of suitable solvents
189 and readily form a film by the solvent casting method.

190 Polymer coating was first carried out using 120 µg of PVC in THF to form
191 a 2.4 µm thick coated film. The TD-GC/MS measurements for 5 µL of the
192 phthalate stock mixture solution were then performed using the polymer-coated
193 sample cup with changing the waiting time. The observed results are shown
194 in Fig .5 for waiting times of zero (a) and 200 min (b). The peak intensities in
195 both chromatograms are comparable for all phthalates. This result suggests

196 that the PVC-coated sample cup works well to reduce the emission of
197 semi-volatile phthalates such as DMP and DEP. The reduction of emission
198 can be ascribed to the fast diffusion of the phthalate mixture solution into the
199 coated PVC film.

200 Next, the phthalate mixture was repeatedly measured five times with
201 different waiting times from 0 to 200 min at an interval of 50 min each. As a
202 result, a good reproducibility in the peak area is obtained for all phthalates with
203 RSD of less than 3%. In addition to the peaks related to phthalates, however,
204 new peaks assigned to naphthalene (peak A) and biphenyl (peak B) derived
205 from the PVC film, appear in the retention time ranging from 3 to 5 min as
206 shown in Fig. 5. In addition, it is considered that a large amount of hydrogen
207 chloride may be produced due to dissociation of the side chain of PVC when
208 heated up to 320°C according to the reported thermogram of PVC [11].
209 However, these compounds originated from PVC do not affect the analytical
210 results on phthalates that show longer retention times than 5 min.

211 Figures 6 and 7 show chromatograms of the phthalate mixture obtained
212 using PS- and PMMA-coated sample cups, respectively. Polymer coating on
213 the inner surface of the sample cup was done by the same procedure as was
214 used in preparation of the PVC-coated sample cup. In both Figs. 6 and 7, the
215 upper chromatograms (a) were obtained immediately after putting the phthalate
216 mixture into the sample cup, and the lower chromatograms (b) were obtained
217 after holding the sample cup containing the phthalate mixture at the waiting
218 position of the Auto-Shot Sampler for 200 min. Irrespective of the holding
219 time for the sample cup, chromatograms are coincident with each other as
220 shown in Figs. 6 and 7. Besides, additional peaks are observed; peaks C
221 (styrene dimer) and D (styrene trimer) in Fig. 6, peaks E (MMA dimers) and F
222 (MMA trimers) in Figs. 7, which are considered to originate from each polymer
223 film. These peaks, however, do not affect the analytical results of the
224 phthalates, since they are well separated from the phthalate peaks. Thus,

225 both PS- and PMMA-coated sample cups can also be used to reduce the
226 emission of semi-volatile phthalates and they are effective for the TD-GC/MS
227 analysis of phthalates including DMP and DEP, the same as the findings for the
228 PVC-coated sample cup.

229

230 3.3 Effect of the amount of a coating polymer on the chromatographic peak
231 area

232 In the former experiment, 120 μg of polymer was used to prepare
233 polymer-coated sample cups for the analysis of phthalates by TD-GC/MS.
234 Here, dependence of the amount of one of the coated polymers (PVC) on the
235 peak areas of phthalates was examined. The obtained results for the mixed
236 sample of 10 phthalates (100 ng each) with the 200 min waiting time are shown
237 in Fig. 8. The less-volatile phthalates show almost constant peak areas
238 irrespective of the PVC amount as shown in the curves of DPRP, DIBP, and
239 DEHP.

240 On the other hand, peak areas of semi-volatile phthalates such as DMP
241 and DEP increase with an increase in the PVC amount, and then reach an
242 almost constant level when the PVC amount is larger than 60 μg . In addition,
243 a good reproducibility with RSD of 2.23 and 2.37% was obtained for the peak
244 areas of DMP and DEP acquired from five sequential measurements with the 60
245 μg PVC-coated sample cup and different waiting times from 0 to 200 min at an
246 interval of 50 min each. Accordingly, it can be said that the PVC-coated
247 sample cup with 60 μg or more of PVC reduces the emission of semi-volatile
248 phthalates and it is applicable to the analysis of phthalates containing DMP and
249 DEP.

250

251 4. Conclusion

252 The polymer-coated sample cups with PVC, PS and PMMA coatings were
253 developed, and their basic performance was evaluated by measuring a mixture
254 of phthalates including semi-volatile DMP and DEP. As a result, no reduction
255 of peak areas of DMP and DEP during the waiting time was observed when
256 using PVC-coated sample cups having film thicknesses of 0.6 μm or more.
257 Comparable performances to quench the emission were also observed using
258 PS- and PMMA-coated sample cups. These results suggest that not only
259 less-volatile phthalates but also semi-volatile ones such as DMP and DEP can
260 be measured with high reliability by TD-GC/MS using the polymer-coated
261 sample cups.

262

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331

332

333 Figure Captions

334 Fig.1 Cross-sectional view of a sample cup coated with a sorbent polymer
335 film.

336 Fig.2 Schematic diagram of the measurement system for TD-GC/MS analysis.

337 Fig.3 Chromatograms of the phthalate mixture measured by TD-GC/MS at up
338 to 320°C using a temperature programmable pyrolyzer obtained with plural
339 different waiting times after putting the phthalate mixture into a sample cup
340 without polymer coating (a) 0 min, (b) 200 min. Sample amount was 100 ng for
341 each phthalate (20 ppm×5 µL). For peak assignments, see Table 1.

342 Fig.4 Effect of the waiting time at 25°C on the peak area of each phthalate
343 obtained by TD-GC/MS using sample cup without polymer coating. ●: DMP, □:
344 DEP, ■: DPRP, ×: DIBP, ○: DEHP.

345 Fig.5 Chromatograms of the phthalate mixture measured by TD-GC/MS at up
346 to 320°C using a sample cup coated with PVC (average film thickness: 2.4 µm)
347 obtained with plural different waiting times; (a) 0 min, (b) 200 min. A:
348 naphthalene, B: biphenyl. For the other peak assignments, see Table 1.

349 Fig.6 Chromatograms of the phthalate mixture measured by TD-GC/MS at up
350 to 320°C using a sample cup coated with PS (average film thickness: 2.4 µm),
351 obtained with plural different waiting times; (a) 0 min, (b) 200 min. C: styrene
352 dimer, D: styrene trimer. For the other peak assignments, see Table 1.

353 Fig.7 Chromatograms of the phthalate mixture measured by TD-GC/MS at up
354 to 320°C using a sample cup coated with PMMA (average film thickness: 2.4
355 µm), obtained with plural different waiting times; (a) 0 min, (b) 200 min. E: MMA
356 dimers, F: MMA trimers. For the other peak assignments, see Table 1.

357 Fig.8 Relationship between the film thickness of PVC and the peak area of
358 phthalates obtained by TD-GC/MS for 200 min waiting time. ●: DMP, □: DEP, ■:
359 DPRP, ×: DIBP, ○: DEHP.

Table 1 Phthalate compounds and their physical properties

Peak No.	Compound name	Abbreviation	MW	BP ^{*1} (°C)	VP ^{*2} (Pa)
1	Dimethyl phthalate	DMP	194	282	2.63×10^{-1}
2	Diethyl phthalate	DEP	222	298	6.48×10^{-2}
3	Di-n-propyl phthalate	DPRP	250	317	1.75×10^{-2}
4	Diisobutyl phthalate	DIBP	278	327	4.73×10^{-3}
5	Di-n-butyl phthalate	DBP	278	340	4.73×10^{-3}
6	Di-n-pentyl phthalate	DPP	306	342	1.28×10^{-3}
7	Di-n-hexyl phthalate	DHP	334	-	3.45×10^{-4}
8	Butyl benzyl phthalate	BBP	312	370	2.49×10^{-3}
9	Di (2-ethylhexyl) phthalate	DEHP	390	386	2.52×10^{-5}
10	Di (n-octyl) phthalate	DNOP	390	380	2.52×10^{-5}

*1: Boiling point, *2: Vapor pressure at 25°C

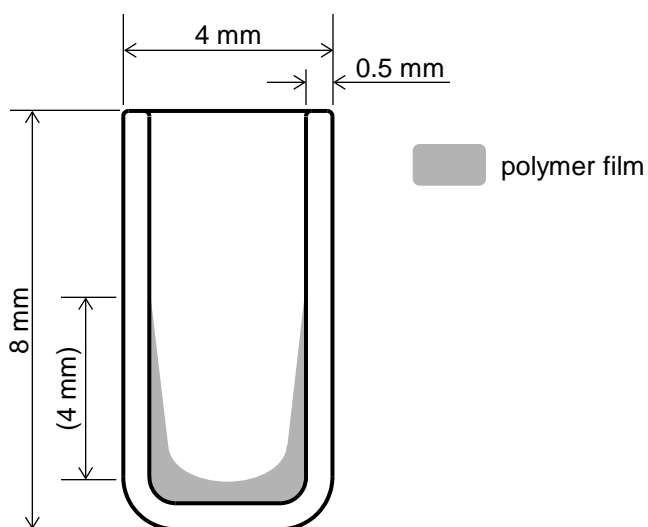


Fig.1 Cross section of a polymer-coated glass sample cup.

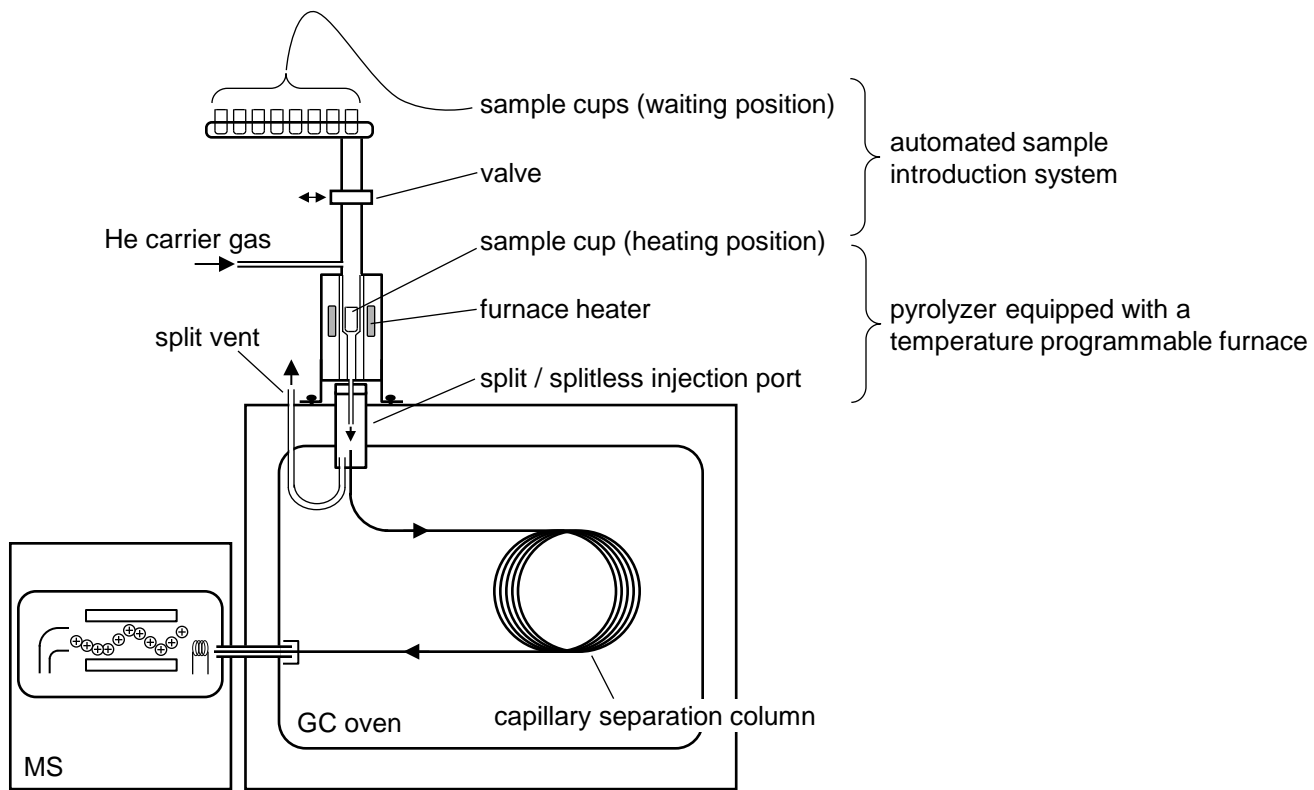


Fig.2 Schematic diagram of the measurement system for TD-GC/MS analysis.

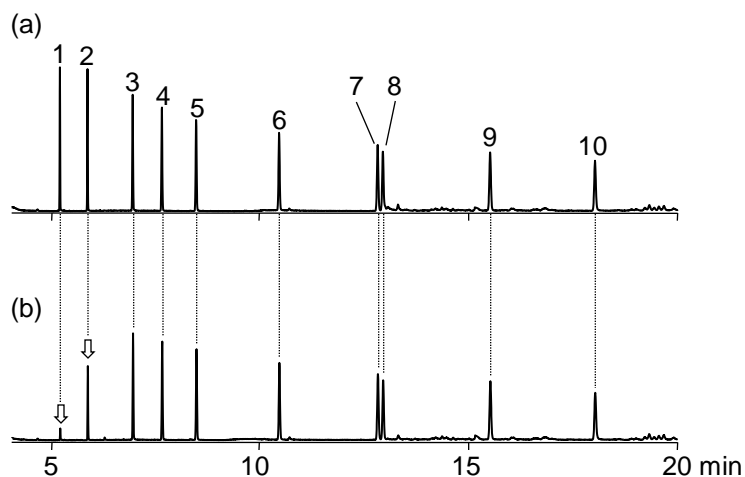


Fig.3 Chromatograms of the phthalate mixture measured by TD-GC/MS at up to 320°C using a temperature programmable pyrolyzer obtained with different waiting times after putting the phthalate mixture into a sample cup without polymer coating (a) 0 min, (b) 200 min. Sample amount was 100 ng for each phthalate (20 ppm \times 5 μ L). For peak assignments, see Table 1.

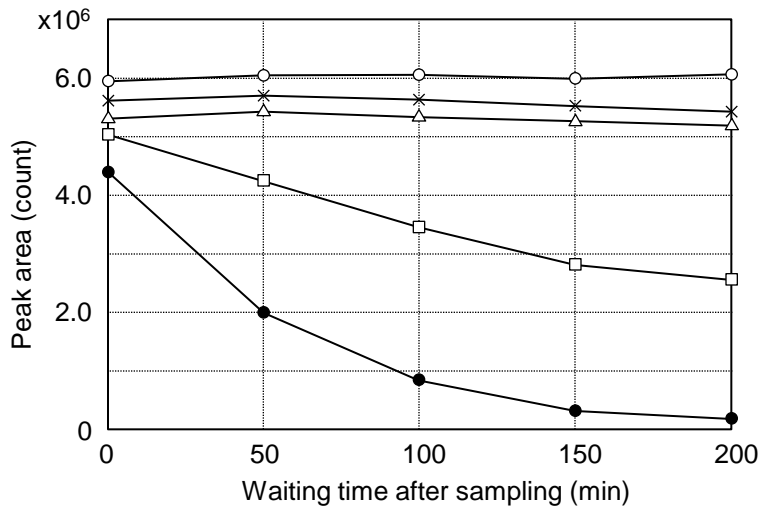


Fig.4 Effect of the waiting time at 25°C on the peak area of each phthalate obtained by TD-GC/MS using sample cup without polymer coating. ●: DMP, □: DEP, △ DPRP, ×: DIBP, ○: DEHP.

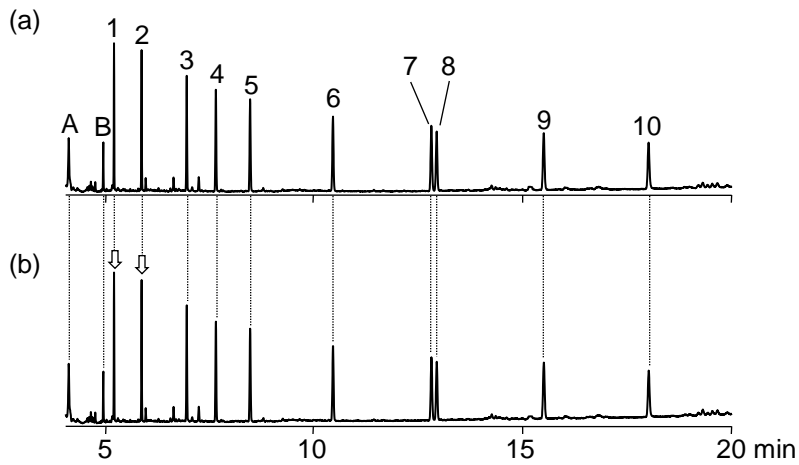


Fig.5 Chromatograms of the phthalate mixture measured by TD-GC/MS at up to 320°C using a sample cup coated with PVC (average film thickness: 2.4 μm) obtained with different waiting times; (a) 0 min, (b) 200 min. A: naphthalene, B: biphenyl. For the other peak assignments, see Table 1.

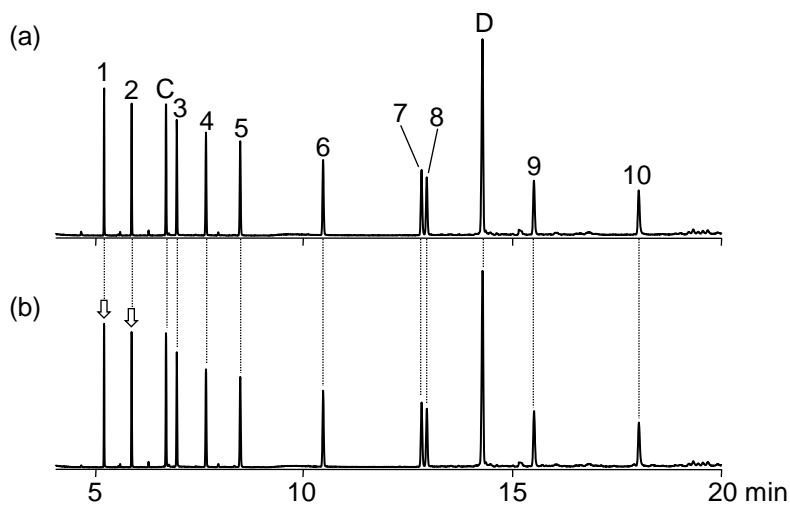


Fig.6 Chromatograms of the phthalate mixture measured by TD-GC/MS at up to 320°C using a sample cup coated with PS (average film thickness: 2.4 μm), obtained with different waiting times; (a) 0 min, (b) 200 min. C: styrene dimer, D: styrene trimer. For the other peak assignments, see Table 1.

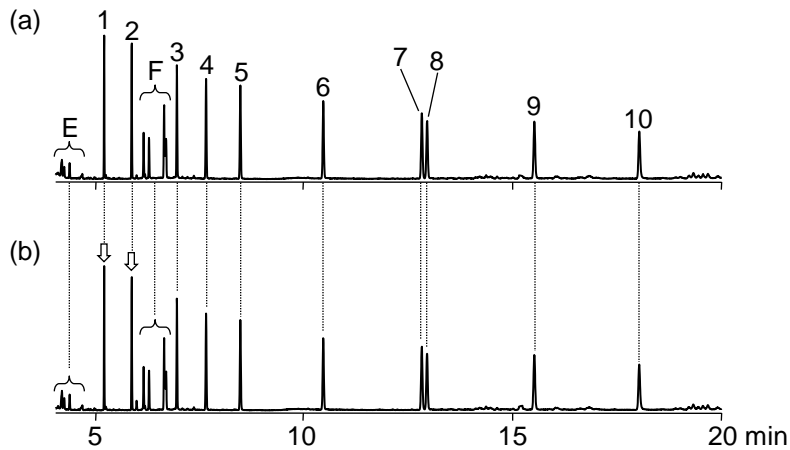


Fig.7 Chromatograms of the phthalate mixture measured by TD-GC/MS at up to 320°C using a sample cup coated with PMMA (average film thickness: 2.4 μm), obtained with different waiting times; (a) 0 min, (b) 200 min. E: MMA dimers, F: MMA trimers. For the other peak assignments, see Table 1.

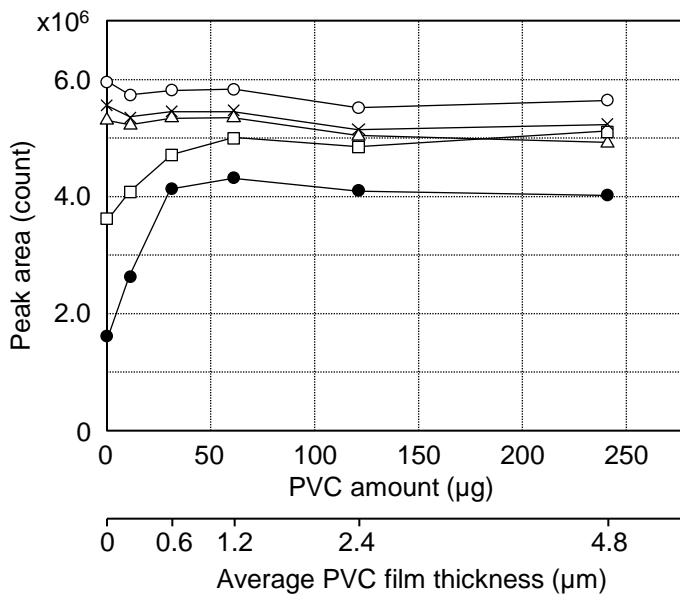


Fig.8 Relationship between the film thickness of PVC and the peak area of phthalates obtained by TD-GC/MS for 200 min waiting time. ●: DMP, □: DEP, △: DPRP, ×: DIBP, ○: DEHP.